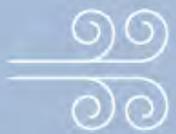


operates in different fields and turns
Ultraviolet Technology into real
Solutions, providing a Specific Device for
every application needed.



HVAC



Water



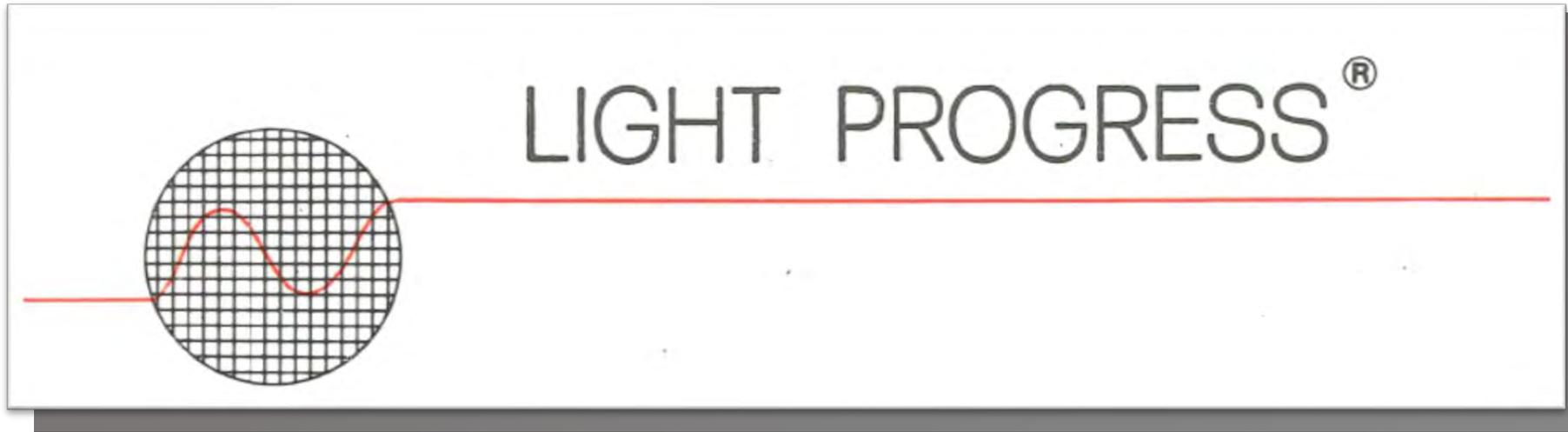
Health



Food



Smell reduction



studies, develops, projects and produces

Ultraviolet Germicidal Irradiation

devices, since



UltraViolet Germicidal Irradiation

What does UVGI mean?

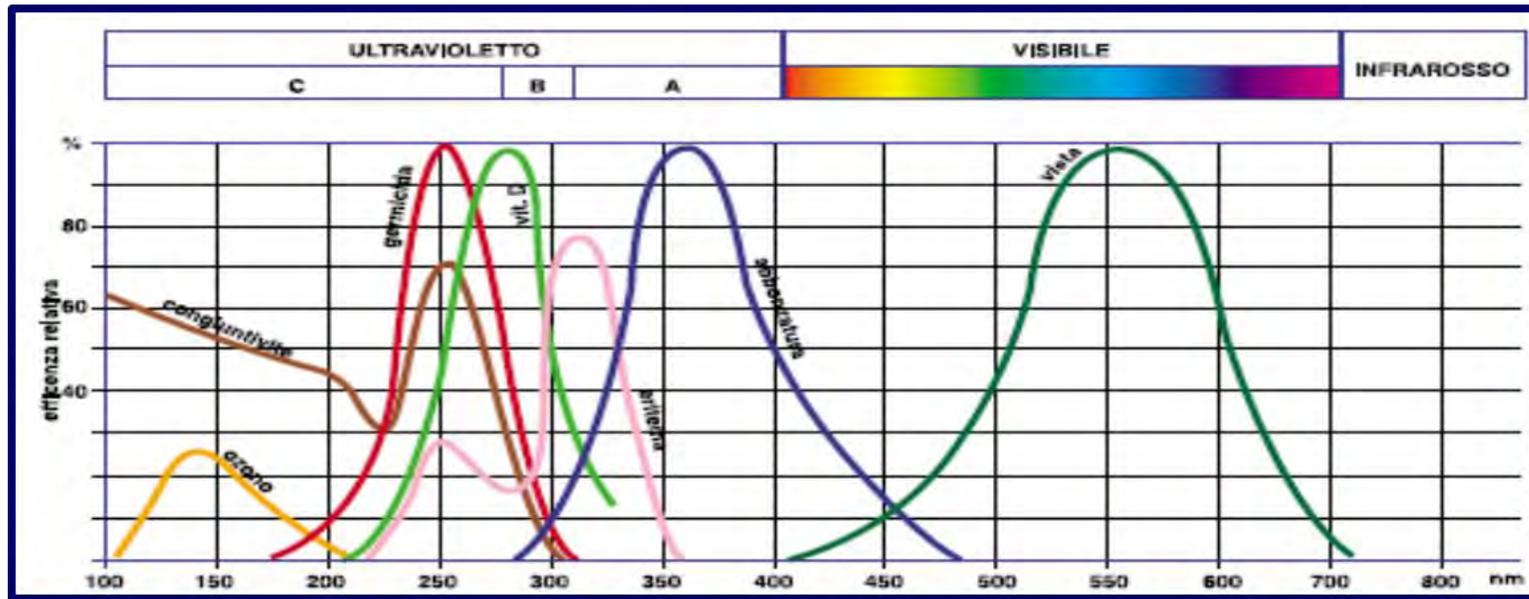
Light in a broad sense can be divided in visible, infra-red and ultraviolet rays.

Ultra-violet rays (invisible) can be classified in:

- UV - A (with tanning properties)
- UV - B (with therapeutic properties)
- **UV - C (with germicidal properties)**



Ultraviolet Germicidal Irradiation is known from the 60s as a good physical method to control **growth and distribution of microbial organisms, pathogens, spores, moulds, etc.**



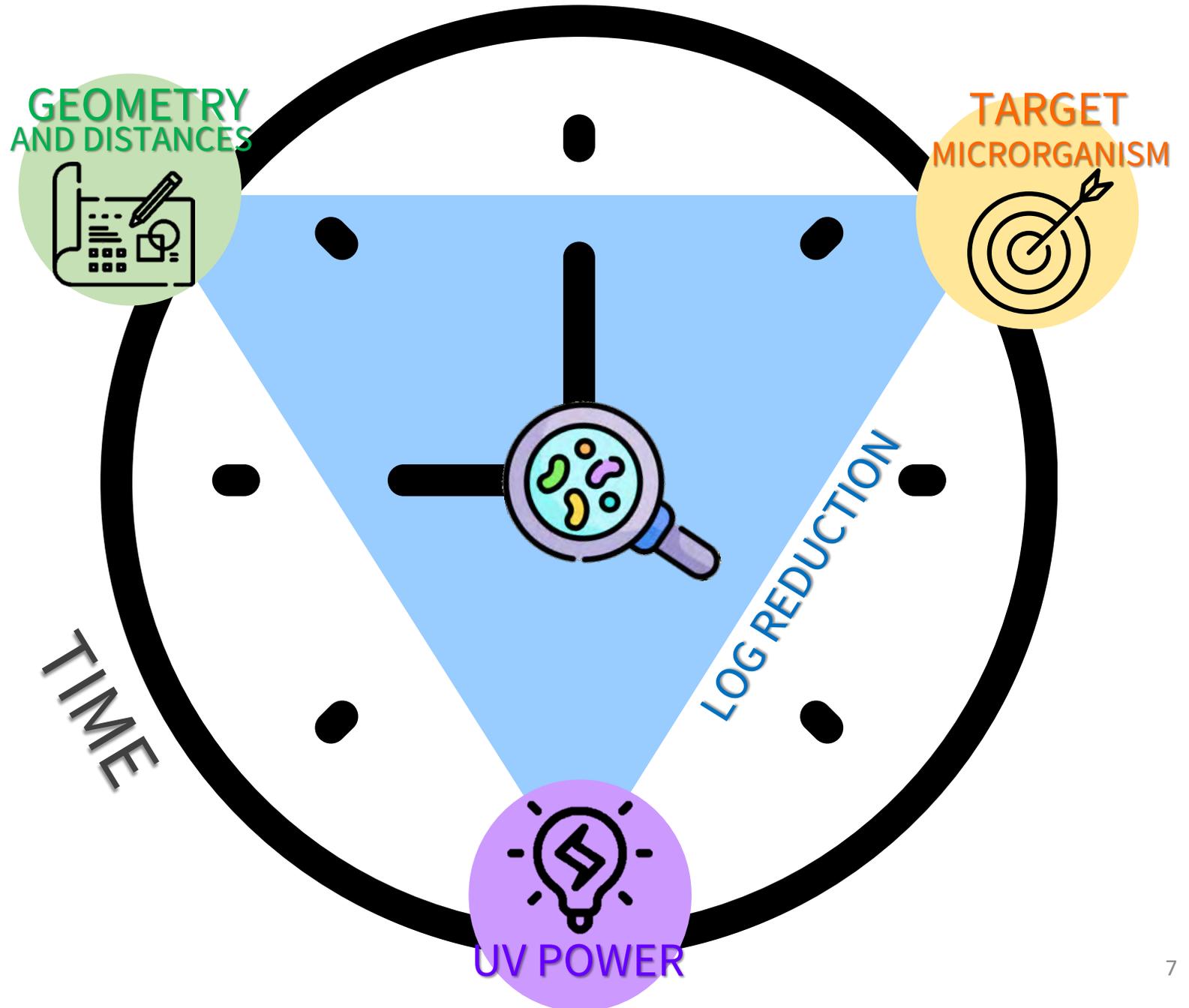
UV-C (short-wave) from 100 to 280 nm

The absorption of a UV photon by the DNA of microorganisms causes a destruction of a link in the DNA chain, and consequently the inhibition of DNA replication.

The germicidal effects of the UV-C radiation **destroy DNA of Bacteria, Viruses, Spores, Fungi, Moulds and Mites** avoiding their growth and proliferation.

UVGI technology is a physic disinfection method with a **great costs/benefits** ratio, it's ecological, and, unlike chemicals, it works against every microorganisms without creating any resistance.

UV Disinfection Key Factors



SIMPLIFIED CALCULATION OF THE DOSES TO ACHIEVE KILLING OF 90% OF COMMON MICROORGANISMS

**NOTE: FOR DESTRUCTIONS OF 99% TO 99.9% - 99.99% OF MICROORGANISMS (AND SO ON),
THE DOSES SHOULD BE INCREASED RESPECTIVELY 2 - 3 - 4 TIMES**

MICROORGANISM	DOSE (µW/cm2 SEC)	MICROORGANISM	DOSE (µW/cm2 SEC)	MICROORGANISM	DOSE (µW/cm2 SEC)
BACTERIA		BACTERIA		RNA VIRUS	
Bacillus (vegetative)	3200 (1300-5800)	Proteus vulgaris	2.600		
B. anthracis	4.500	Pseudomonas	3500 (1500-5500)	Picomavirus	7200 (3600-18600)
B. subtilis	5.800	Pseudomonas aeruginosa	5.500	Poliovirus	11.000
B. megatherium	1.300	Pseudomonas fluorescens	3.500	Poliov type 1 Mahoney	6.700
B. paratyphosus	3.200	Salmonella	4300(2100-8000)	Poliov	13.300
Bacillus (spore)	11800 (1100-36500)	Salmonella enteritis (gastroenterite)	4.000	Poliov type 1	3.600
B. Megatherium	2.700	S. typhosa (febbre tifoidea)	4.000	Poliov Mahoney	4.500
B. subtilis	12.000	S. paratyphi (febbre enterica)	3.200	ECBO	8.000
Bacillus anthracis	4.500	S. typhimurium	8.000	Coxsackiev	18.600
B. subtilis spore ATCC6633	15.200	Sarcina lutea	19.700	Reovirus	10200 (4800-16300)
Campylobacter jejuni	2.900	Serratia marcescens	2.400	Reovirus type 1	4.800
Clostridium tetani	13.000	Shigella	1700 (1700-4400)	Reovirus type 1 (Lang. Str.)	16.300
Clorynebact. diphtheria	3.400	Shigella dysenteriae (diarrea)	4.200	Rotavirus	15.900
Citrob. Freundii (ATCC8090)	4.200	Shigella flexneri (diarrea)	1.700	Rotavirus SA11	6.500
Enterobacterium cloaca (ATCC13047)	6.400	Shigella paradysenteriae	1.700	Paramyxovirus	3500 (1500-5500)
Eberthella typhosa	2.100	Staphylococcus	4400 (1800-11000)	Sindbis Virus	5.500
Escherichia coli	4500 (700-5800)	Staphylococcus albus	1.800	Newcastle Disease	1.500
Escherichia coli	3.000	Staphylococcus aureus	2.600	HIV (Lentiv)	143800 (60000-240000)
Escherichia coli (in air)	700	Staphylococcus epidermis	11.000	HIV (HTLVIII)	60.000
Escherichia coli (in water)	5.400	Streptococcus	3600 (1800-6500)	HIV (Sup. T1)	145.000
Escherichia coli ATCC11229	2.500	Streptococcus hemolyticus	2.200	HIV (H9)	240.000
Escherichia coli ATCC 25922	3.000	Streptococcus lactis	6.200	HIV (PHA- stim. PBL)	130.000
Escherichia coli K 12 AB 1157	5.800	Streptococcus viridans	2.000		
Escherichia coli B/r ATCC 12407	5.300	Streptococcus faecalis (ATCC29212)	6.500	YEAST	
Klebsi. Pneumon. ATCC4352	4.200	Streptococcus faecalis	5.500	Comuni lieviti dolciari	6.000
Legionella	1500 (400-2600)	Streptococcus pyogenes	2.200	Saccharomyces ellips. (panificazione)	6.000
Legionella dumoffi	2.400	Streptococcus salivaris	2.000	Saccharomyces cerevisiae (panificaz.)	6.000
Legionella gormanii	2.600	Streptococcus albus	1.800	Torula sphaerica	2.300
Legionella micdadei	1.500	Mycobacterium tubercoli	10.000	Lievito di birra	10.000
Legionella longbeachae 1	1.200	Vibrio comma (Colera)	3400-6500	SPORES OF MOULDS	
Legionella longbeachae 2	1.000	Yersinia enterocolitica	1.500	Aspergillus amstelodami (carne)	66.700
Legionella oakridgensis	2.200	VIRUS (GEN.)		Aspergillus flavus (verde / giallastro)	60.000
Legionella jordanis	1.100			Aspergillus glaucus (verde / bluastro)	44.000
Legionella wadsworthii	400	Batteriofagi	2.600	Aspergillus niger (panifici) (nero)	132.000
Legionella pneumophila	2.500	Virus dell' influenza	6.000	Cladosporium herbarum (celle frigo)	60.000
Legionella bozemanii	2.000	Virus della poliomelite	6.500	Mucor mucedo (carne, pane, formaggio)	65.000
Leptospira	2000(800-2800)	Virus dell' epatite A	8.000	Mucor racemosus (A e B) (grigio/bianco)	17.000
Leptospira biflexa	2.300	DNA VIRUS		Oospora lactis (bianca)	5.000
Leptospira illini	800			Penicillium digitatum (verde oliva)	44.000
Leptospira interrogans	2.800	Parvovirus	3500 (3000-4000)	Penicillium chrysogenum (frutta)	50.000
Leptospira Spp.- Infectious jaundice	3.200	Bov. parvovirus	4.000	Penicillium roqueforti (verde)	13.000
Leptospira hemorrhagie (Weil sindr.)	2.000	Kilham rat virus	4.000	Penicillium expansum (verde oliva)	13.000
Listeria monocytogenes	9.000	HCC (Dog hepat. Adenov)	26.500	Rhizopus nigricans (nero)	111.000
Micrococcus	8000 (6100-10000)	Herpes virus	5700 (1500-16500)	Rhizopus nigericans (formaggio)	300.000
Micrococcus candidus	6.100	Pseudorabies virus	7.000	Scopulriopsis brevicaulis (formaggio)	80.000
Micrococcus piltonencis	8.100	Herpes simplex MP str.	6.700	ALGAE	
Micrococcus sphaeroides	10.000	Herpes simplex, type 1	16.500	Diatoms, green and blue algae	600.000
Mycobacterium tuberculosis (TBC)	6.200	Pseudomonas aeruginosa	5.500	PROTOZOA	
Neisseria catarrhalis	4.400	Vaccinia	1.800	Paramecium	100.000
Phytomonas tumefaciens	4.400			WORMS	
				Eggs of Nematodes	40.000

SANITATION means bringing the microbial load into acceptable and optimal hygiene standards that depend on the intended use of the environments concerned. Sanitation is often used to mean “clean” and must however be preceded by cleaning.

SANITATION

DISINFECT means to reduce the microbial load deeply, that is to eliminate at least 1 log (90%) of the bacteria present. Microbial load reduction is a basic value in disinfection and it is expressed in Log Reduction.

A good disinfection level is around 2Logs (99%) a very good disinfection is 3Logs (99,9%), and 4Logs (99,99%) is considered a pretty high standard.

DISINFECTION

STERILITY is the closest level anyone can get to achieve complete reduction of microbial load, we can talk about sterilization only if reduction is proved to be not less than 6Logs, meaning 99,9999%.

To declare sterility test has to be proved and certified by third parts by law.

STERILIZATION

Why LIGHT PROGRESS ?

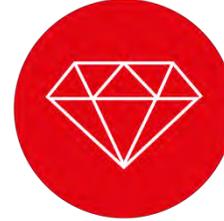
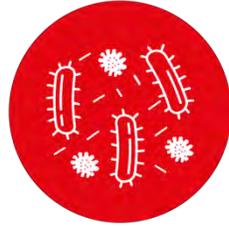
} We offer the **widest product range** of UVGI Devices on the market, providing different solutions, specifically designed for any application.

} Our Team has the knowledge to consider all the different key factors and the combination between them. With this know-how we size every application achieving **the targets solution** for each specific case.

} Our products are born answering **real market issues coming from different Industries**, our first mission is to fit UV-C in already existing environments, satisfying exactly the clients requests.

Benefits

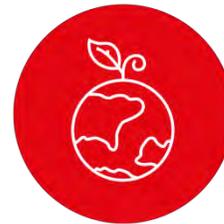
We eliminate every harmful microorganism, up to 99,99%



We improve Quality



We ensure you safety



We support sustainability



We make you save money



Our team is there to support you



Certificates



LIGHT PROGRESS

CE

DECLARATION OF COMPLIANCE

We, LIGHT PROGRESS S.r.l., hereby declare under our own responsibility that the following units of own production:

[Redacted]

- ⇒ are in accordance with EEC directive 2014/30/EU (Electromagnetic Compatibility)
- ⇒ are in accordance with EEC Machinery Directive dispositions 2006/42/EU
- ⇒ are in accordance with EEC Low Voltage Directive 2014/35/EU
- ⇒ are in accordance with EEC (RoHS) directive 2002/95/EU and 2011/65/EU

TECHNICAL STANDARDS APPLIED

UNI EN ISO 12100:2010 Safety of Machinery - Basic Concepts, General Principles for Design - Risk assessment and risk reduction

UNI EN ISO 13857:2008 Safety of Machinery - Safety Distances to prevent danger zones being reached by the upper and lower limbs (2008)

ISO 14120:2015 Safety of Machinery - Guards - General Requirements for the Design and construction of fixed and movable guards

UNI EN ISO 13849-1:2016 Safety of Machinery - Parts of the Control System related to the Safety - Part 1: General Design Principles

UNI EN ISO 14119:2013 Safety of Machinery - Interlocking devices associated with guards - Principles for design and selection

CEI EN 60204-1:EC Safety of Machinery - Electrical Equipment of Machines. Part 1: General Rules (2010)

EN 61439-1:2011 Low-voltage Switchgear and Control Gear Assemblies. Part 1: General rules

FURTHER TECHNICAL STANDARDS APPLIED:

IEC EN 60335-1 "Safety of household electrical appliances and similar"
Electronic Ballasts for the control of the lamps in accordance with the standard EN 60928.
Germicidal UV-C Lamps in accordance with EN 61199.
Electrical Protective Measures in accordance with IEC 70-1, EN 60229.

Anghiari, 05 January 2017

Responsible for Standards: Dr. Aldo Santi

LIGHT PROGRESS S.r.l. Via G. Marconi, 81 - 53031 ANGIARI (AR) - ITALY - <http://www.lightprogress.com>

Jan-2017 Pag. 22/24

kiwa
Partner for progress

Reg. Number	6950 - A	Valid From	2016-07-26
First issue date	2007-12-21	Last modification date	2013-07-24
Following renewal date	2019-07-29	EA Sector	EA: 19

Quality Management System Certificate
ISO 9001:2015

We certify that the Quality Management System of the Organization:
LIGHT PROGRESS S.r.l.

is in compliance with the standard UNI EN ISO 9001:2015 for the following products/services:

Design and production of UV-C rays disinfection systems

Chief Operating Officer
Giampiero Balcredi

Maintenance of the certification is subject to annual survey and dependent upon the observance of Kiwa Cermet Italia contractual requirements.

This certificate consists of 1 page.

LIGHT PROGRESS S.r.l.
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Certified sites
- Località San Antonino 40 52043 Castiglion Fiorentino (AR) Italia
- Via Guglielmo Marconi, 81 52031 Anghiari (AR) Italia

CERMET **IAF** **ACCREDIA** ISO 9001:2015 ISO 9001:2008 ISO 9001:2015 FISN N° 0241 PRD N° 0698

CERTIFICATE OF COMPLIANCE*

Certificate Number	20130702-E362672
Report Reference	E362672-20130628
Issue Date	2013-JULY-02

Issued to: LIGHT PROGRESS SRL
Via G. MARCONI 81
52031 ANGIARI AR ITALY

This is to certify that representative samples of ACCESSORIES, AIR-DUCT MOUNTED Duct-Mounted UV Lamp Assembly, Models UV-RACK, followed by 3I, 4I or 6I, followed by 40H, 60H or 90H.

Have been investigated by UL in accordance with the Standard(s) indicated on this Certificate.

Standard(s) for Safety: Bi-National Standard for Heating and Cooling Equipment, ANSI/UL 1995-2011 and CAN-CSA C22.2 No. 236-11

Additional Information: See the UL Online Certifications Directory at www.ul.com/database for additional information

Only those products bearing the UL Classification Mark for the U.S. and Canada should be considered as being covered by UL's Classification and Follow-Up Service and meeting the appropriate U.S. and Canadian requirements.

The UL Classification Mark includes: the UL in a circle symbol: with the word "CLASSIFIED" (as shown), a control number (may be alphanumeric) assigned by UL; a statement to indicate the extent of UL's evaluation of the product; and the product category name (product identity) as indicated in the appropriate UL Directory. The UL Classification Mark for Canada includes: the UL Classification Mark for Canada: with the word "CLASSIFIED" (as shown), a control number (may be alphanumeric) assigned by UL; a statement to indicate the extent of UL's evaluation of the product; and the product category name (product identity) in English, French, or English/French as indicated in the appropriate UL Directory.

Look for the UL Classification Mark on the product.

William R. Casey, Director, North American Certification Programs
UL LLC

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Page 1 of 1

*Available for selected items



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University Tests - Air Treatment



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Valutazione dell'effetto che purificatori d'aria a raggi UV-C prodotti da **LIGHT PROGRESS®** hanno sulla carica microbica e fungina presente nell'aria.

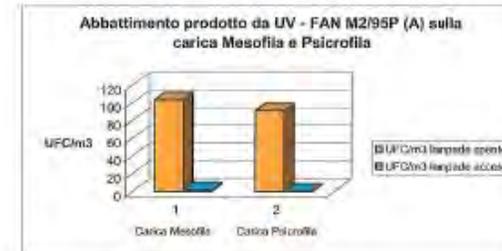
University of Siena
Department of Physiopathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Meroneci

Emanuele Meroneci
Stefano Biondi

Grafico 1



Grafico 2



University of Siena
Department of Physiopathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Meroneci

Em



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University Tests - Microbial Load Reduction



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Valutazione dell'effetto battericida, sporicida e fungicida dei raggi UV-C emessi da apparecchi LIGHT PROGRESS®

University of Siena
Department of Pathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Montanelli

Emanuele Montanelli

Aspergillus niger



Aspergillus niger su Sabouraud dextrose Agar, a sinistra la piastra non irradiata, a destra la piastra irradiata con UVC.

University of Siena
Department of Pathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Montanelli

EM

Escherichia coli



Escherichia coli su MacConkey Agar n.3, a sinistra la piastra non irradiata, a destra la piastra irradiata con UVC.

University of Siena
Department of Pathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Montanelli

EM

Staphylococcus aureus



Staphylococcus aureus su Mannitol salt agar, a sinistra la piastra non irradiata, a destra la piastra irradiata con UVC.

University of Siena
Department of Pathology,
Experimental Medicine and Public Health
Lab. Molecular Epidemiology
Prof. Emanuele Montanelli

EM

Gráfico 1

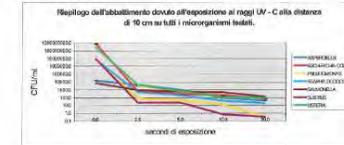
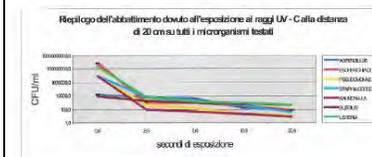


Gráfico 2



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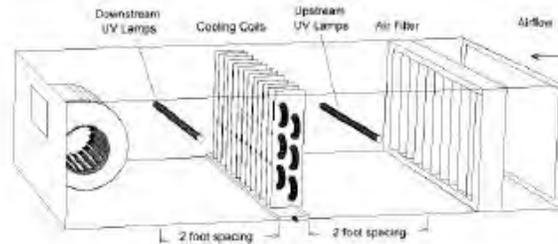


Figure 2.1: Location and spacing for UVGI system in an air handling unit

2.2 Location of UV Lamp Ballasts

UV lamp ballasts should preferably be located external to the ventilation system although this is not currently a strict requirement due to so many systems that have integral lamp ballasts that must be located wherever the lamp is located. One of the problems with lamp ballasts being located inside air handling units is that they may be exposed to temperature and humidity extremes.

If lamp ballasts are located in an internal lamp housing, the housing should be of drip-proof construction or other approved housing method. If lamp ballasts are located external to the air handling unit or ductwork, the wiring must be run through conduit such that there is no exposed wiring inside the air handling unit. Exposed wiring may be subject to deterioration inside and air handling unit and may also be exposed to UV irradiation, which may cause photodegradation over time and thus pose a fire hazard.

2.3 Operating Conditions

Both the UV lamp and the ballast should be located such that the ambient operating conditions (i.e. temperature and relative humidity) are within the component design or operating limits. Refer to manufacturer's information for design operating conditions. In general, both UVGI and filters are designed to operate at an air velocity of 500 fpm, although some lamps may be suitable for operation at higher velocities. Variations in air velocity (i.e. +/- 100 fpm) may be acceptable depending on the manufacturer's lamp but such variations should be evaluated to include or assess the impact on UV output. See IJVA-G01A-2005, "General Guideline for UVGI Air and Surface Disinfection Systems," for

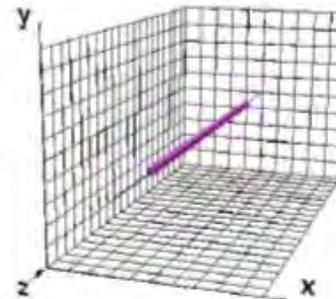


Figure 6.1: Grid for a 16x16x20 Matrix and Coordinate System, shown with a lamp in an axial configuration.

6.2 Operation of the Program

The program takes the input data from an input text file, performs the analysis and outputs results in a text file. Intermediate results can be extracted and graphed in spreadsheets.

Input data requires definition of the coordinate system. The lamp coordinates are based on the lower left front corner of the matrix being at (0, 0, 0). The indices for both the large and small matrices are also based on this (0, 0, 0) point.

Using the input the enclosure intensity field is determined by evaluating the direct intensity field of the lamp, the first reflection intensity field, and the total inter-reflected intensity field. These are summed to produce the total intensity field of the enclosure. This process is shown by the flow chart in Figure 6.2.

As mentioned previously, the inter-reflectors are only computed for three iterations, after which the total bulk average intensity is determined mathematically for the remaining inter-reflections. Each of the first three inter-reflection calculations involves computing the exchange of radiative energy from each of the blocks on the other three sides, for all four walls. The summed result produces the wall intensity contours for the next set of inter-reflection calculations. This is the most calculation-intensive portion of the program and takes up the most operating time. In comparison, the direct and first reflection calculations proceed relatively rapidly.

Because two different size matrices are used for the computations, it is necessary to scale up the smaller matrix to match the size of the larger matrix prior to adding them. This is

Effective UVGI System Design Through Improved Modeling

W.J. Kowalski, P.E.
Student Member ASHRAE

William P. Bahnfleth, Ph.D., P.E.
Member ASHRAE

ABSTRACT

This paper summarizes an improved methodology for predicting the actual ultraviolet disinfection for UVGI systems that will usually effective design and lower energy costs. This approach uses radiative transfer theory to design the three-dimensional intensity field for lamps and reflective surfaces inside enclosures. Lamp photometric data for a variety of lamps are shown to agree more closely with the inverse square model than with models using the inverse square law. The intensity field due to reflection from internal surfaces is determined by assuming diffuse reflectivity. An analytical method is used to determine the inter-reflection component of intensity due to multiple internal reflections. The superposition of these components yields a three-dimensional intensity field source that can be used to calculate disinfection rates for any given microbial rate constant. Results from laboratory bioassays using *S. aureus* in various duct configurations have corroborated model predictions within ±10% in most cases.

INTRODUCTION

Currently available design information has not guaranteed predictable performance for UVGI air disinfection systems. Some of today's design practices can overdesign systems, leading to prohibitive costs and high energy consumption. Other design practices lead to underdesigned and ineffective systems. Design practices have not changed in decades, and it is worthwhile to review the history of UVGI applications to determine how this situation has come to be.

Although the first UVGI water disinfection system was implemented in 1909 (AN/NA 1971), the first UVGI systems designed for maximum disinfection were implemented until the 1930s (Shapp 1949). Based on limited laboratory data and

using rarely available UVGI lamps, these systems were sized within the benefits of positioning remote tests, either as sampling or endpoints point, were used to determine their efficacy. Some of these systems were highly successful, such as those used to control measles in schools, and one used by Riley to eliminate TB bacilli from hospital ward surfaces in (Riley and O'Grady 1961).

Other designs appeared to be ineffective, with the result that the actual glowing reviews of this technology became fragmented. Guidelines were issued that sanctioned the use of UVGI only in combination with HEPA filters (Luciano 1977, ASHRAE 1981). No studies were ever undertaken to determine the root cause for any UVGI system failures. Apart from improvements in lamp design, applications technology for maximum disinfection has remained almost stagnant for decades.

The first design guidelines for UVGI air disinfection systems were developed in the 1940s (Lackock and Holladay 1942; Lackock 1946). Versions appeared in catalogs that continue to be reproduced and used today (Phillips 1993). These guidelines offer procedures, charts, and tables to size lamps and reflective surfaces so as to obtain a desired disinfection rate. These sizing methods, though intuitively detailed for the period, suffer from a number of deficiencies:

1. They did not define the intensity field, instead merely using the lamp rating or relying on photometric data for lamp application.
2. Lamps are specified without regard to lamp location or type.
3. The correction factor for rectangular duct ignores the intensity field variations due to surface reflectivity.

W.J. Kowalski is a doctoral candidate and William P. Bahnfleth is an associate professor in the Department of Architectural Engineering, Pennsylvania State University, University Park, Pa.

Thank you

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